Heavy Metals Present in Industrial Effluents and Their Effect on Fish: An Overview

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Abstract- This review gives a brief account of the toxic effects of heavy metals on fish. In aquatic ecosystem, heavy metals are considered as the most important pollutants, since they are present throughout the ecosystem and are detectable in critical amounts. Heavy metals, such as cadmium, copper, lead, Nickle and zinc are of the most pollutants which important effect environment and fish. They are extremely hazardous for the health of fish. Most of these heavy metals are characterized by being accumulated in tissues, and lead to the poisoning of fish. These metals can effectively influence the vital operations and reproduction of fish; weaken the immune system, and induce pathological changes. As such, fish are used as bio-indictors, playing an important role in monitoring heavy metals pollution.

Indexed Terms- Aquatic pollution, Bio-indicators, Fish, Heavy metals,

I. INTRODUCTION

At present, the pollution has become a serious threat, and has brought hazards to the growing population as well as the earth environment. The contamination of fresh waters with a wide series of pollutants has become a matter of concern over the last few years (Voegborlo et al., 1999; Canli et al., 1998). The rapid urbanization and industrialization has led to increased disposal of pollutants like heavy metals, radio nuclides, and various types of organic and inorganic substances into the environment. The metal which has a relatively high density and toxic at low quantity is referred as 'heavy metal', e.g., arsenic (As), lead (Pb), mercury (Hg), cadmium (Cd), chromium (Cr), thallium (Tl), etc. Some 'trace elements' are also known as heavy metals, e.g., copper (Cu), selenium (Se) and zinc (Zn). They are essential to maintain the body metabolism, but they are toxic at higher concentrations. The heavy metals can enter the bodies

to a small extent via food, drinking water and air. The heavy metals concerned with the environmental science chiefly include Pb, Hg, Cd, Cr, Cu, Zn, manganese (Mn), nickel (Ni), silver (Ag), etc. Further, the heavy metals are metallic elements which have a relatively high density, and they are poisonous at low quantity. The excess quantities of heavy metals are detrimental as these destabilize the ecosystems because of their bioaccumulation in organisms, and elicit toxic effects on biota and even death in most living organisms (Gupta, 2013).

The industrial wastes are the main source of metal pollution for aquatic organisms. It has been cited that the heavy metals constitute the major pollutants in the environment. The paper industry has been one of the chief industries causing aquatic pollution because the effluent releasing by it is highly toxic. The pulp and paper mill industry is one of the oldest industries in India and there has been tremendous expansion of these industries during last 25 years. Many chemicals have been identified in effluents which are produced at different stages of papermaking. Their toxic nature is derived from the presence of several naturally occurring and xenobiotic compounds which are formed and released during various stages of papermaking (Sangeeta *et al.*, 2013).

The heavy metals are important pollutants for fishes, because these are not eliminated from aquatic systems by natural methods, such as organic pollutants, and are enriched in mineral organic substances. The metal contaminants are mixed in the aquatic system through smelting process, effluents, sewage and leaching of garbage which cause severe harm to the aquatic system.

The main problem with heavy metals in our bodies is their ability to bio-accumulate. Bio-accumulation means the metals do not leave the body by their own accord and accumulate in certain tissues. Due to bioaccumulation, heavy metals are passed up the food chain from smaller species (fish) to humans. The main tissues targeted by heavy metals include: the liver, kidneys, bowel, brain and nervous system, spleen and eyes (Sharma and Agrawal, 2005).

The natural aquatic systems may extensively be contaminated with heavy metals released from domestic, industrial and other man-made activities. Heavy metal contamination may have devastating effects on the ecological balance of the recipient environment and a diversity of aquatic organisms (Farombi, *et al.*, 2007; Vosyliene and Jankaite, 2006; Ashraj, 2005).

Among animal species, fishes are the inhabitants that cannot escape from the detrimental effects of these pollutants (Olaifa et al., 2004; Clarkson, 1998; Dickman and Leung, 1998). Fish are widely used to evaluate the health of aquatic ecosystems because pollutants build up in the food chain and are responsible for adverse effects and death in the aquatic systems (Farkas et al., 2002; Yousuf and El-Shahawi, 1999). The studies carried out on various fishes have shown that heavy metals may alter the physiological activities and biochemical parameters both in tissues and in blood (Basa and Rani, 2003; Tort and Torres, 1988). The toxic effects of heavy metals have been reviewed, including bioaccumulation (Wagar 2006; Adami et al., 2002; Rasmussen and Anderson, 2000; Rani, 2000; Aucoin et al., 1999).

Even though some of the heavy metals such as zinc, iron, cobalt and copper are essential for enzymatic activity and other biological processes at low levels they become toxic when they exceed certain limit. On the other hand other metals such as lead, cadmium and mercury have no essential role in living organisms and are toxic even at too low concentrations (Bryan, 1976). Heavy metals do not pend in water and settle down swiftly onto sediment due to their higher density than that of water. This was demonstrated with Cd and Cu exposure, metals showed 72 to 97% decrease from their initial concentration after 96 hours of experiment (Ghosal and Kaviraj, 2002; Ghosh *et al.*, 2016; Ghosh *et al.*, 2018).

The bioaccumulation of heavy metals in different tissues of aquatic organisms leads to several harmful effects. It may have genotoxic, mutagenic, Immunosuppressive and cytotoxic effects. It may also result in histopathological changes, abnormalities in fish reproduction; and public health hazard effects for human consumption such as polluted fish.

II. CLASSIFICATION OF HEAVY METALS

- Essential heavy metals Copper, chromium, zincnickel, cobalt and iron are essential metals required for all vital processes inside the body with optimum level. Otherwise, inadequate amount causes deficiency diseases and high-level cause's toxicity (Sivaperumal *et al.*, 2007).
- 2. Non-essential heavy metals- Those haven't biological roles and also called xenobiotic. When they are increased in concentrations, it will cause toxopathic effects in tissue; those involve Aluminum, Mercury, Lead, Cadmium and others. (Sfakianakis *et al.*, 2015).

• Cadmium (Cd):

Cadmium is the seventh most toxic heavy metal as per ATSDR ranking. Cadmium distributed in the environment will remain in soils and sediments for several decades. Plants gradually take up these metals which get accumulated in them and concentrate along the food chain, reaching ultimately the human body. Once this metal gets absorbed by humans, it will accumulate inside the body throughout life. Fish exposed to cadmium revealed a negative effect on their the growth rate, meat quality and blood and biochemical parameters (Vijayram et.al 1989; Shukla et al., 2002; Abbas et al., 2007). The morphological and histological alterations in the liver, intestine and kidneys of cadmium exposed fish (Thophon et al., 2003; Omer et al., 2012). Higher doses of cadmium caused visible external lesions such as discoloration of fish (Cavas et al., 2005). The cadmium poisoning caused softening of the bones and kidney failure. It can cause osteoporosis, nonhypertrophic emphysema, irreversible renal tubular injury, eosinophilia, anosmia and chronic rhinitis. Chronic exposure to inorganic Cd results in accumulation of the metal mainly in the liver and kidneys as well as in other tissues and organs causing many metabolic and histological changes, membrane damage, altered gene expression and apoptosis (Soeginato 2008; Bais and Lokhande, 2012).

• Chromium (Cr):

Chromium is a metal found in natural deposits such as rocks (ores), animals, plants and soil and can be a liquid, solid or gas Chromium is used in textile industries, electroplating, leather tanning, metal finishing and chromate preparation, protective coatings on metal (electroplating), magnetic tapes and pigments for paints, cement, paper, rubber, composition floor covering and other materials. Its soluble forms are used in wood preservatives. Chromium compounds bind to soil and are not likely to migrate to ground water, but they are very persistent in sediments in water. Like other heavy metals, chromium enters the body of fish through gills or digestive tracts, but it is known that it has a lower ability to accumulate than others (Rashed, 2001). The accumulation of Cr (VI) in the kidney led to macroscopic and microscopic abnormalities and negatively affected fish growth and survival (Farag et al., 2006). Impact of chromium trivalent and hexavalent toxicity on the behavior of Danio rerio (Zebra fish), erratic motion, mucus discharge, opening mouth for gasping, color and shade alteration, irregular swimming was usually point out (Nisha, 2016). The behavioral changes in gold fish (Carrassius arratus) and noted that all the fingerlings come to the aquarium and there was also appetite decrease due to chemical effects (Faward, et al., 2017). Acute poisoning by chromium compounds causes excess mucous secretion, damage in the gill respiratory epithelium and the fish may die with symptoms of suffocation (Benoit ,1976).

Chromium (VI) compounds are toxic and known human carcinogens, whereas chromium (III) is less toxic and an essential nutrient for human (WHO, 1988; Jordao *et al.*, 2002; Abbas *et al.*, 2016; Martin and Griswold, 2009). Exposure to higher amounts of chromium compounds in humans can lead to the inhibition of erythrocyte glutathione reductase, which in turn lowers the capacity to reduce methemoglobin to hemoglobin (Koutras *et al.*, 1965).

• Copper (Cu):

Copper (Cu) is an essential trace metal and micronutrient for cellular metabolism in living organisms on account of being a key constituent of metabolic enzymes. However, it can be extremely toxic to intracellular mechanisms in aquatic animals at

high concentrations which exceed normal levels (Hernández *et al.*, 2006). Although the crucial role of Cu in several enzymatic processes, this heavy metal can exert adverse toxicological effects, when present in high concentrations in water. In fact, it is potentially toxic when the internal available concentration exceeds the capacity of physiological detoxification processes. Increasing agricultural production has resulted in increasing number of freshwater systems being impacted by the contaminants present in wastewater releases (Li, and Wu, 1999).

It is an abundant element which occurs as a natural mineral with a wide spread use. Copper pollution is through extensive use of fungicides, algaecides, molluscicides, insecticides and discharge of industrial wastes. Copper sulfate (CuSO4) is often used as an algaecide in commercial and recreational fish ponds to control the growth of phytoplankton and filamentous algae and to control certain fish disease. Fish can accumulate copper via diet or ambient exposure (Sfakianakis et al., 2015). Even at low environmental concentrations, copper shows distinct affinity to accumulate in the fish liver. It decreases the total protein content in liver and muscles with increasing the concentration of free amino acids gluconeogenic enzyme activity in copper exposed fish. The typical patho-anatomical appearance includes a large amount of mucus on body surface, under the gill covers and in between gill filaments (Svobodova ,1993). Cu exposed different fish species posed behavioural changes such as decrease in swimming ability and food intake and increase in operculum movements (Venkataramana and Radhakrishnaiah, 2001; Ali et al., 2003). Hepatocyte vacuolization, necrosis, shrinkage, nuclear pyknosis and increase of sinusoidal spaces were the distinct changes observed in the liver of copper-exposed fish (Figueiredo-Fernandes et al., 2007). Gainey and Kenyon (1990) mentioned that exposure of fishes to sublethal concentrations of copper leads to cardiac activity and reduction in heart rate. Radhakrishnaiah et al. (1992) have recorded stimulation of glycogenolysis in fish L. rohita on exposure to a sub lethal concentration of copper. Sanchez et al. (2005) showed that Cu is able to induce oxidative stress in fish (Gasterosteus aculeatus) even before significant metal accumulation occurs in the liver.

Copper plays an important role in human metabolism, largely because it allows many fundamental and essential enzymes to function properly and also plays a role in the production of hemoglobin, myelin, and melanin. Copper is playing an important role as transition metal signaling, transferring information in and beyond the brain, between and within the living cells. Some significant evidences were found that a physiological imbalance of the redox-active bio metals, Cu and Fe, and oxidative stress lead to the neuropathology of Alzheimer's disease. The Menkes syndrome and Wilson disease are also due to the severe copper deficiency and severe copper toxicity, respectively (Turnlund, 2005).

• Lead (Pb):

Lead (Pb) is a naturally occurring substance, its environmental concentrations are significantly increased by anthropogenic sources which include base metal mining, battery manufacturing, Pb-based paints and leaded gasoline. Lead in water may come from industrial and smelter discharges; from the dissolution of old lead plumbing, lead containing pesticides, through precipitation, fallout of lead dust, street runoff, and municipal wastewater (Sepe, *et al.*, 2003).

The main mechanisms of lead toxicity are the activation of cellular functions due to this metal's calcium mimicking effect, and the inhibition of the activity of different proteins through its binding to sulfhydryl groups. Lead has high affinity for sulfhydryl groups and can inactivate enzymes, especially those involved in heme synthesis, such as aminolevulinic acid dehydratsase and ferrochelatase (Gwalteney-Brant, 2002). Fish exposed to 5 ppm of lead nitrate for 150 days, it exhibited marked inhibition of gonadal growth and showed decrease in cholesterol and lipid levels in brain, testis and ovary whereas the liver showed an elevation of both (Katti, et al., 1983).

Acute lead toxicity is initially characterized by damaging gill epithelium and ultimately suffocation. Two types of structural alterations of gill, defense/compensatory responses and direct deleterious effects were observed in chronic lead exposed fish (Parashar and Banerjee, (1999). The necrosis and desquamation of gill epithelium as well as lamellar curling and

aneurisms were the direct deleterious effects reported in chronic lead exposed *Clarias gariepinus* (Olojo 2005). The characteristic symptoms of chronic lead toxicity include changes in the blood parameters with severe damage to erythrocytes and leucocytes and damage in the nervous system (El-Badawi, (2005). Low levels of Pb pollution could cause some adverse effects on fish health and reproduction (Delistraty and Stone, 2007). Also, lead was found to inhibit the impulse conductivity by inhibiting the activities of mono aminooxidase and acetylcholine esterase, to cause pathological changes in tissue and organs (Rubio,1991).

Chronic lead poisoning has similar toxic effects in fish as in mammals. These include hematological and neural disorders and tetanic spasms together with some morphological changes such as darkening in caudal fin, deformation of vertebrate, anomalies in pigment formation and covering of the gills by a mucus layer (Shah, 2006).

• Nickel (Ni):

Nickel is a non-biodegradable toxic heavy metal ion present in waste water. The main source of nickel pollution in the water derives from a number of industrial production processes such as battery manufacturing, production of some alloys, zinc base casting, printing, electroplating and silver refineries(Li et al., 2000; Meche et al., 2010). Once released to the environment, nickel readily forms complexes with many ligands, making it more mobile than most heavy metals .While nickel is an essential element at low concentrations for many organisms, it is toxic at higher concentrations (Palaniappan and Karthikeyan, 2009).

Primarily, Nickel accumulate in the gills, kidney and muscles of fishes. Nickle toxicity is responsible for increased mucus secretion, dark red colour of gill, to swell up the gill lamellae as well as increasing oxygen consumption, ventilatory stroke volume and respiratory rate (Sreedevi et al., 1992; Pane et al., 2004; Ghosh et al., 2018). Nickel alters the haematological parameters (erythrocyte, leucocytes, hematocrit and hemoglobin count) and lowered values of mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC) in fishes when compared with the control (Al-Ghanim ,2011).

• Zinc (Zn):

Zinc (Zn) is the second most abundant trace element after Fe and is an essential trace element and micronutrient in living organisms, found almost in every cell and being involved in nucleic acid synthesis and occurs in many enzymes. Zinc is an essential trace element for both plants and animals but it induces toxicity at elevated concentrations particularly under conditions of low pH, low alkalinity, low dissolved oxygen and elevated temperatures (Eisler, 2010).

Zinc dissolves in water, and releases free zinc ions which become primary source of aquatic toxicity (Blinova et al., 2010). Additionally, Zn is involved in more complicated functions, such as the immune system, neurotransmission and cell signaling (Celik, 2004). Cicik, (2003) also suggested that high concentrations of Zn might have occured due to mucus secretion and structural alterations in gill tissue caused by contamination. Zinc causes morphological changes, alteration in haematological and biochemical parameters, mortality, growth retardation, respiratory and cardiac changes, inhibition of spawning, and a multitude of additional detrimental effects which threaten survival of fish. Gill, liver, kidney, and skeletal muscle are damaged (Sorensen ,1991; Srivastava and Prakash, 2018a, b&c).

III. IMPACT OF HEAVY METAL ON HUMAN HEALTH

Every water body receives the effluents containing heavy metals either from point or from nonpoint sources. These heavy metals persist in environment due to their unbiodegradable nature and accumulate in the tissues of aquatic fauna particularly fish, and thus, they remain in the tissues of the fish for long time. Fishes are the important source of protein. Unfortunately, fishes are now becoming the major source of heavy metals due to the pollution caused by industries. Health problems caused by low level chronic exposure to heavy metals may take years to appear. According to United States Environmental Protection Agency (USEPA), these metals generally cause two types of health effects. One is carcinogenic and other is non-carcinogenic effects. These heavy metals have been linked to conditions ranging from cardiovascular disease, high blood pressure, insomnia, and many more. Much of the exposure to heavy metal

pollution has been scientifically proven to be linked to causing free radical damage leading to: Heart attacks, strokes and cancer. There are also many circulatory diseases other than cardiovascular disease that may not cause death, but can affect the quality of life. Some of these are: Impotency, Alzheimer's disease, Arthritis, Diabetes, Fatigue, Memory Loss. Lead poisoning is a serious and very common type of heavy metal poisoning. Symptoms of lead poisoning in children mimic those of attention deficit hyperactivity disorder (ADHD). Lead poisoning also causes behavioral and learning problems, nervousness, headaches, and many other related symptoms. Pregnant women, lactating mother and children are more prone to heavy metal health hazards.

Lead poisoning in children causes neurological damage leading to a reduction in intelligence, loss of short-term memory, learning disabilities and problems with coordination. Prenatal exposure can cause reduced birth weight and immune suppression or oversensitisation, which could explain why some children develop asthma and allergies (Day, 1998).

It has been detected that Pb inhibits Na+/K+-ATPase enzyme and d-aminolevulinic acid dehydratase enzyme that participates in growth and hem synthesis in erythrocytes and affects lipid peroxidation enzyme. It has been shown that Pb also has influence on intercellular communication by changing alanine aminotransferase (ALT) and aspartate aminotransferase (AST) concentrations in tissues and organs (Cogun and Şahin, 2012).

CONCLUSION

Based on the present review it is believed that for biomonitoring studies only single parameter is insufficient. Hence, biomarkers such as bioaccumulation, blood profiles, biochemical profiles, pathological marker enzyme activities, enzymatic and non-enzymatic antioxidants, lipid peroxidation, DNA damage and tissue damage could serve as useful tools to monitor the health of aquatic fauna. These heavy metals will enter the food web through water and food, to cause the adverse health effects like that in indicator organisms (fish) as well as their consumers.

REFERENCECS

- [1] Abbas,A.,AlAmer,A.M.,Laoui,T.,AlMarri,M.J., Nasser,M.S.,Khraisheh,M.andAtieh,M.A.(2016) . "Heavy metal removal from aqueous solution by advanced carbon nanotubes: critical review of adsorption applications", *Sep Purif Technol. Elsevier B.V*,Vol.157,141-161.
- [2] Abbas HHH, Hammada M and Miller JD (2007) Vitamin C and cadmium toxicity in fish Oreochromis niloticus. *Online J. Vet. Res.*, 11(1): 54-74.
- [3] Adami, G.M., P. Barbieri, M. Fabiani, S. Piselli, S. Predonzani, E. and Reisenhofer, (2002). Levels of cadmium and zinc in hepatopancreas of reared *Mytilus galloprovincialis* from the Gulf of Trieste (Italy). *Chemosphere*, 48(7): 671-677.
- [4] Al-Ghanim KA (2011) Impact of nickel (Ni) on hematological parameters and behavioral changes in *Cyprinus carpio* (common carp). *Afr J Biotechnol* 10: 13860-13866.
- [5] Ali, A., Al-Ogaily, S.M., Al-Asgah, N.A. and Gropp, J. (2003). Effect of sublethal concentrations of copper on the growth performance of *Oreochromis niloticus*. *J.Appl. Ichthyol*. 19: 183-188.
- [6] Ashraj, W. (2005). Accumulation of heavy metals in kidney and heart tissues of *Epinephelus microdon* fish from the Arabian Gulf. *Environ. Monit. Assess.*, 101(1-3): 311-316.
- [7] Aucoin, J., R. Blanchand, C. and Billiot,(1999). Trace metals in fish and sediments from lake Boeuf, South Eastern Louisiana. *Micro. Chem. J.*, 62(2): 299-307.
- [8] Bais UE and Lokhande MV, (2012), Effect of cadmium chloride on histopaqthological changes in the fresh water fish Ophiocephalus stratus (Channa). *International Journal of Zoological Research*, 8(1), 23-32.
- [9] Basha P.S. and A.U. Rani, (2003). Cadmium induced antioxidant defense mechanism in freshwater teleost *Oreochromis mossambicus* (Tilapia). *Ecotoxicol. and Environ. Safety.*, 56(2): 218-221.

- [10] Benoit DA (1976). Toxic effects of hexavalent chromium on brook trout (Salvelinus fontinalis) and rainbow trout (Salmo gairdneri). *Water Res*, 10: 497-500.
- [11] Blinova, I., Ivask, A., Heinlaan, M., Mortimer, M., Kahru, A. (2010). Ecotoxicity of nanoparticles of CuO and ZnO in natural water. *Environ Pollut*. 158:41–7.
- [12] Bryan, G.W. (1976). Some aspects of heavy metal tolerance in aquatic organisms. In: A.P.M. Lockwood (Ed.), Effects of Pollutants on Aquatic organisms. Cambridge University Press., UK: 7-34.
- [13] Canli, M., O. Ay, M. Kalay, (1998). Levels of heavy metals (Cd, Pb, Cu, and Ni) in tissue of *Cyprinus Carpio*, *Barbus Capito* and *Chondrostoma regium* from the Seyhan river. Turk. J. Zool., 22(3): 149-157.
- [14] Cavas T, Garanko NN, and Arkhipchuk VV (2005). Induction of micronuclei and binuclei in blood, gill and liver cells of fishes subchronically exposed to cadmium chloride and copper sulphate. *Food Chem Toxicol* 43: 569-574.
- [15] Celik U, Oehlenschläger J (2004). Determination of zinc and copper in fish samples collected from Northeast Atlantic by DPSAV. *Food Chem* 87: 343347.
- [16] Cicik, B. (2003). Bakır-çinko etkileşiminin sazan (Cyprinus carpio)'nın karaciğer, solungaç ve kas dokularındaki metal birikimi üzerine etkileri. Ekoloji Çevre Dergisi, Cilt 12, Sayı 48: 32-36.
- [17] Clarkson, T.W., (1998). Human toxicology of mercury. J. Trace. Elem. Exp. Med., 11(2-3): 303-317.
- [18] Çoğun, H.Y., Şahin, M. (2012). Nil Tilapia (Oreochromis niloticus Linnaeus, 1758)'da Kurşun Toksisitesinin Azaltılmasında Zeolitin Etkisi. Kafkas Univ Vet Fak Derg. 18(1): 135-140.
- [19] Day M. (1998). Lead in the womb. *New Scientist*, 23 May, p. 7.
- [20] Delistraty, D. and Stone, A. (2007). Dioxins, metals, and fish toxicity in ash residue from

- space heaters burning used motor oil. *Chemosphere*, 68(5): 907-914.
- [21] Dickman, M.D. and K.M. Leung, (1998). Mercury and organo chlorine exposure from fish consumption in Hong Kong. *Chemosphere*, 37(5): 991-1015.
- [22] Eisler, R. (2010) Compendium of trace metals and marine biota 2. Amsterdam: Vertebrates. *Elsevier*; 2010.
- [23] El-Badawi AA (2005) Effect of lead toxicity on some physiological aspects of Nile tilapia fish, *Oreochromis niloticus*. *Inter. Conf. Vet. Res. Div.*, 2005, NRC, Cairo, Egypt.
- [24] Farag AM, May T, Marty GD, Easton M, Harper DD, Little EE, Cleveland L (2006): The effect of chronic chromium exposure on the health of Chinook salmon (*Oncorhynchus tshawytscha*). *Aquatic Toxicology*, 76, 246–257.
- [25] Farkas, A., J. Salanki, A. Specziar, (2002). Relation between growth and the heavy metal concentration in organs of bream *Abramis brama* L. populating lake Balaton. *Arch. Environ. Contam. Toxicol.*, 43(2): 236-243.
- [26] Farombi, E.O. O.A. Adelowo, Y.R. Ajimoko., (2007). Biomarkers of oxidative stress and heavy metal levels as indicators of environmental pollution in African Cat fish (*Clarias gariepinus*) from Nigeria Ogun river. *Int. J. Environ. Res. Public Health.*, 4(2): 158-165.
- [27] Faward M. Yousafzai A M, Haseeb A. Rehman U H, Afridi A J, Akhtar N. (2017). Acute toxicity and bioaccumulation of chromium in gills skin and intestine of gold fish (Carassius auratus), *Journal of entomology and zoology*. 5(1):568-571.
- [28] Figueiredo-Fernandes A, Ferreira-Cardoso JV, Garcia-Santos S, Monteiro SM, Carrola J, Matos P, et al. (2007): Histopathological changes in liver and gill epithelium of Nile tilapia, *Oreochromis niloticus*, exposed to waterborne copper. *Pesq. Vet. Bras* 27: 103-109.
- [29] Gainey LF and Kenyon JR. (1990). The effects of reserpine on copper induced cardiac inhibition in

- Mytilus edulis. Comp. Biochem. Physiol. C 95: 177-179.
- [30] Ghosal, T.K. and Kaviraj, A. (2002). Combined effects of cadmium and composted manure to aquatic organisms. *Chemosphere*. 46(7): 1099–1105.
- [31] Ghosh, A., Kaviraj, A. and Saha, S.(2016). Kinetics of deposition, acute toxicity and bioaccumulation of copper in some freshwater organisms. *Bull Environ Contam Toxicol*. 97: 820–825.
- [32] Ghosh, A., Kaviraj, A. and Saha, S. (2018). Deposition, acute toxicity, and bioaccumulation of nickel in some freshwater organisms with best-fit functions modeling. *Environ Sci Pollut Res*. 25: 3588–3595.
- [33] Gupta, V.(2013). Mammalian Feces as Bio-Indicator of Heavy Metal Contamination in Bikaner Zoological Garden, Rajasthan, India, Res. J. Animal, Veterinary and Fishery Sci., 1(5), 10-15.
- [34] Gwalteney-Brant, S.M(.(2002). Heavy metals. In: Haschek, W.M., Rosseaux, C.G., Wallig, A.M. (Eds.), *Handbook of Toxicologic Pathology*. Academic Press, New York, 701–732.
- [35] Hernández PP, Moreno V, Olivari FA, Allende ML (2006) Sub-lethal concentrations of waterborne copper are toxic to lateral line neuromasts in zebrafish (*Danio rerio*). *Hear Res* 213: 1-10.
- [36] Jordao, C., Pereira, M., & Pereira, J. (2002). Metal contamination of river waters and sediments from effluents of kaolin processing in Brazil. *Water, Air, & Soil Pollution*, Vol. 140, No. 1, pp 119-138, ISSN 0049-6979 Kamal.
- [37] Katti SR, Sathyanesan AG (1983). Lead nitrate induced changes in lipid and cholesterol levels in the freshwater fish *Clarias batrachus*. *Toxicol*. *Lett*. 19: 93-96.
- [38] Koutras GA, Schneider AS, Hattori M, Valentine WN. (1965). Studies on chromated erythrocytes. Mechanisms of chromate inhibition of glutathione reductase. *Br J Haematol* 11(3): 360–369.

- [39] Li, L. and Wu, G.(1999). Numerical Simulation of Transport of Four Heavy Metals in Kaolinite Clay. *J Environ Engineering*, 125(4); 314-324.
- [40] Li, X., Shen, Z., Wai, O.W.H., Li, Y., Li, X. and Shen, Z. (2000). "Chemical partitioning of heavy metal contaminants in sediments of the pearl river estuary", *Chemical Speciation and Bioavailability*, Vol.12No.1,: 17-25.
- [41] Martin, S.,Griswold,W.(2009)."Human health effects of heavy metals", Cent Hazard Subst.1-6.
- [42] Meche, A., Martins, M.C., Lofrano, B., Hardaway, C.J., Merchant, M. and Verdade, L. (2010). "Determination of heavy metals by inductively coupled plasma-optical emission spectrometry in fish from the Piracicaba river in Southern Brazil", *Microchemical Journal*, Vol. 94 No. 2.171-174.
- [43] Nisha JC, Sekar RRJ, Chandran R,(2016) Acute effect of chromium toxicity on the behavioral response of Zebra fish Danio rerio. International Journal of plant "Animal and environmental sciences. 6(2) 6-14.
- [44] Olaifa, F.G., A.K. Olaifa, T.E. Onwude, (2004). Lethal and sublethal effects of copper to the African Cat fish (*Clarias gariepnus*). *Afr. J. Biomed. Res.*, 7: 65-70.
- [45] Olojo EAA, Olurin KB, Mbaka G, Oluwemimo AD (2005) Histopathology of the gill and liver tissues of the African catfish, *Clarias gariepinus* exposed to lead. *Afr. J. Biotechnol.* 4: 117-122.
- [46] Omer SA, Elobeid MA, Fouad D, Daghestani MH, Al-Olayan EM, et al. (2012). Cadmium bioaccumulation and toxicity in tilapia fish (*Oreochromis niloticus*). *J Anim Vet Adv* 11: 1601-1606.
- [47] Palaniappan PLRM and Karthikeyan S (2009). Bioaccumulation and depuration of chromium in the selected organs and whole body tissues of freshwater fish Cirrhinus mrigala individually and in binary solutions with nickel. *J Environ Sci* 21: 229-236.
- [48] Pane, E.F., Richards, J.G., Wood, C.M., (2004). Acute waterborne nickel toxicity in the rainbow trout (*Oncorhynchus mykiss*) occurs by a

- respiratory rather than an iono regulatory mechanism. *Aquat. Toxicol.* 63: 65-82.
- [49] Parashar RS and Banerjee TK (1999) Histopathological analysis of sublethal toxicity induced by lead nitrate to the accessory respiratory organs of the airbreathing teleost, *Heteropneustes fossilis* (Bloch). *Pol. Arch. Hydrobiol.* 46: 199-206.
- [50] Radhakrishnaiah K, Venkataramana P, Suresh A, Sivaramakrishna B (1992) Effects of lethal and sublethal concentrations of copper on glycolysis in liver and muscle of the freshwater teleost, *Labeo rohita* (Hamilton). *J. Environ. Biol* 13: 63-68.
- [51] Rani, A.U.,(2000). Cadmium induced bioaccumulation in tissue of freshwater teleost Oreochromis mossambicus. Ann. N.Y. Acad., 919(1): 318-320.
- [52] RASHED MN.(2001). Monitoring of environmental heavy metals in fish from Nasser Lake. *Environ Int*, 27: 27-33. DOI: 10.1016/S0160-4120 (01)00050-2.
- [53] Rasmussen, A.D., O. Anderso, (2000). Effects on cadmium exposure on volume regulation in the lugworm, Arenicola marina. *Aquat. Toxicol.*, 48: 151-164. Gulf of Trieste (Italy). *Chemosphere*, 48(7): 671-677.
- [54] Rubio R, Tineo P, Torreblanca A, Del Ramo J, Diaz Mayans J (1991). Histological and electron microscopical observations on the effects of lead on gills and midgut gland of Procambarus clarkii. *Toxicol Environ Chem* 31: 347-352.
- [55] Sanchez W, Palluel O, Meunier L, Coquery M, Porcher JM, Aït-Aïssa S (2005). Copper-induced oxidative stress in three-spine stickleback: relationship with hepatic metal levels. Environ. *Toxicol. Pharmacol* 19: 177-183.
- [56] Sangeeta Dey, Manabendra Dutta Choudhury and Suchismita Das. (2013) A Review on Toxicity of Paper Mill Effluent on Fish. Bull. Env. Pharmacol. Life Sci. 2(3): 17-23.
- [57] Sepe A, Ciaralli L, Ciprotti M, Giordano R, Fumari E, Costantini S (2003). Determination of cadmium, chromium, lead and vanadium in six

- fish species from the Adriatic Sea. Food Addit. *Contam.* 20: 543-552.
- [58] Sfakianakis DG, Renieri E, Kentouri M, Tsatsakis AM (2015). Effect of heavy metals on fish larvae deformities: A review. *Enviro. Res.*, 137: 246-255.
- [59] Shah, S.L.2006. Hematological parameters in tench Tinca tinca after short term exposure to lead, *Journal of Applied Toxicology*, 26; 223.
- [60] Sharma RK, Agrawal M (2005). Biological Effects of Heavy Metals: An Overview. *J. Environ Biol. 2005 Jun.*; 26(2 suppl): 301-13.
- [61] Shukla V, Rathi P, Sastry KV (2002). Effect of cadmium individually and in combination with other metals on the nutritive value of fresh water fish, *Channa punctatus*. *J Environ Biol* 23: 105-110.
- [62] Sivaperumal P, Sankar T V, Nair V. 2007. Heavy metal concentration in fish, shellfish and fish products from internal markets of India vis-à-vis international standards Food Chem,102:612-620. DOI: 10.1016/j. foodchem.2006.05.041.
- [63] Soeginato A, (2008). Bioaccumulation of heavy metals in some commercial animals caught from selected coastal waters of East Java, Indonesia. *Research Journal of Agriculture and Biological Sciences*, 4(6), 881-885.
- [64] Sorensen EMB (1991). Metal poisoning in fish: Environmental and Life Sciences Associates. Boca Raton: CRC Press Inc.
- [65] Sreedevi P, Suresh A, Sivaramakrishna B, Prabhavathi B, Radhakrishnaiah K (1992) .Bioaccumulation of nickel in the organs of the freshwater fish, Cyprinus carpio, and the freshwater mussel, *Lamellidens marginalis*, under lethal and sublethal nickel stress. *Chemosphere* 24: 29-36.
- [66] Srivastava, N. K. and Prakash, S (2018a). Morphological, Behavioural and Haematological alterations in catfish, *Clarias batrachus* (Linn.) after Acute Zinc Toxicity. *International journal* on Biological Sciences. 9(1): 72-78.

- [67] Srivastava, N. K. and Prakash, S (2018b). Effect of Sublethal concentration of zinc sulphate on the serum biochemical parameters of freshwater cat fish, *Clarias batrachus*. *Indian Journal of Biology*, 5(2): 113-119.
- [68] Srivastava, N. K. and Prakash, S (2018c). Alterations in the organic reserve of zinc exposed fresh water catfish, *Clarias batrachus*. *Flora and Fauna*, 24(2):386-392.
- [69] Svobodová Z (1993). Water Quality and Fish Health. FAO, Rome, EIFAC technical paper No. 54, 67.
- [70] Thophon, S., M. Kruatrachue, E.S. Upatham, P. Pokethitiyook, S. Sahaphong and S. Jaritkhuan, (2003). Histopathological alterations of white seabass, *Lates calcalifer*, in acute and subchronic cadmium exposure. *Environmental Pollution*, 121: 307-320.
- [71] Tort, L., Torres, P., Hidalgo, J. (1988). The effects of sublethal concentrations of cadmium on haematological parameters in the dogfish Scyliorhinus canicula. J. Fish Biol. 32: 277-282.
- [72] Turnlund, J.R., Keyes, W.R., Kim, S.K. and Domek, J.M. (2005). Long-Term High Copper Intake: Effects on Copper Absorption, Retention, and Homeostasis in Men. American Journal Clinical Nutrition,81,822-828.
- [73] Venkataramana, P., Radhakrishnaiah, K. (2001).
 Copper influenced changes in lactate dehydrogenase and G-6-PDH activities of freshwater teleost, *Labeo rohita*. Arch. Environ.
 Contam. Toxicol. 67: 247-263.
- [74] Vijayram K, Geraldine P, Varadarajan TS, John G, Lognanthan P (1989). Cadmium induced changes in the biochemistry of an air breathing fish *Anabas testudineus*. *J. Ecobiol.*, 1: 245-251.
- [75] Vosyliene, M.Z., A. Jankaite, (2006). Effect of heavy metal model mixture on rainbow trout biological parameters. *Ekologija.*, 4: 12-17.
- [76] Voegborlo, R.B., A.M.E. Methnani, M.Z. Abedin, (1999). Mercury, cadmium and lead content of canned Tuna fish. *Food Chem.*, 67(4): 341-345.

- [77] Waqar, A., (2006). Levels of selected heavy metals in Tuna fish. *Arab. J. Sci. Eng.*, 31(1A): 89-92.
- [78] Yousuf, M.H.A. and M.S. El-Shahawi, (1999). Trace metals in Lethrinus lentjan fish from Arabian Gulf: Metal accumulation in Kidney and Heart Tissues. *Bull. Environ. Contam. Toxicol.*, 62(3): 293-300.